

AMINOGLYCOSIDE ANTIBIOTICS: SYNTHESIS OF
6-O-(β -D-RIBOFURANOSYL) PAROMAMINE

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To date, all the known microbially-derived 4,5-linked aminoglycoside antibiotics contain a β -D-pentofuranosyl moiety attached to carbon-5 of the deoxy-streptamine portion, as in the paromomycin-neomycin¹-lividomycin², and ribostamycin³-butirosin⁴ groups. In a general program directed toward the synthesis and chemical modification of aminoglycoside antibiotics⁵, it was of interest to prepare the title compound, a positional isomer of a bioactive pseudotrisaccharide^{6,7} and to study the biochemical consequences of the "unnatural" attachment of the D-ribofuranose moiety. Paromamine⁸ was chosen as the aglycon because it lends itself to preferential functionalization. A further advantage is the structural versatility of the anticipated pseudotrisaccharide and its derivatives, since a conversion from the paromamine to the neamine-type can be easily achieved by novel procedures⁹.

Benzoylation of the di-O-isopropylidene acetal of tri-N-benzoyloxycarbonyl paromamine^{5,10} followed by preferential cleavage of the acetal function (80% AcOH) gave the crystalline 3'-benzoate derivative¹¹ 1 in 95% overall yield m.p 247-248° (MeOH); $[\alpha]_D^{20} + 91.6$ (DMF). Treatment of 1 with benzaldehyde in 98% formic acid (25°, 4.5 h), followed by neutralization, and usual processing gave the 4',6'-O-benzylidene acetal 2 as a chromatographically homogeneous amorphous solid (85%), m.p 315° (charring); $[\alpha]_D^{20} + 44.3^\circ$ (dioxane). The glycosidation reaction between 2 and 2,3,5-tri-O-benzoyl- β -D-ribofuranosyl chloride¹² 3 was effected in a 1:3 mixture of DMF and 1,2-dichloroethane in the presence of mercuric cyanide and calcium sulfate (50°, 18h). Conventional processing afforded a crude product that consisted essentially of one component (t.l.c, CCl₄, acetone, 4:1 and toluene,

EtOAc, 3:2). Trituration with isopropyl ether and dichloromethane gave the pseudotrisaccharide derivative 4 as a white amorphous solid (96%) which could be further purified by filtration through a bed of silica gel (CCl₄, acetone, 4:1); m.p 278-279°; $[\alpha]_D^{20} + 26.8$ (dioxane)^{13a}. Removal of the acetal function (aq. AcOH-dioxane, 50°, 4d) followed by debenzoylation (NaOMe, MeOH, CH₂Cl₂) gave the tri-N-benzyloxycarbonyl analog 5 (62% overall), m.p 260-265° (dec); $[\alpha]_D^{20} + 41.4$ (DMF), which was found to be different from the 5-O-substituted positional isomer⁶, thus pinpointing the position of attachment of the D-ribofuranosyl moiety in the glycosidation reaction. Hydrogenolysis in the presence of 20% Pd-C in a mixture of methanol and dioxane containing 1N hydrochloric acid, followed by treatment of the product with Rexyn (OH⁻) resin gave the pseudotrisaccharide 6 as a white solid (77%), m.p 174-175°; (sintering at 145°); $[\alpha]_D^{20} + 34.2^\circ$ (H₂O)^{13b}; t.l.c (silica gel, CHCl₃, MeOH, NH₄OH, 1:3:2, R_f 0.53; R_{paromamine} 1.07); paper chromatography (n-PrOH-pyridine-H₂O-AcOH, 15:10:12:3; R_{paromamine} 1.24). The structure of the product 6 was further substantiated by high resolution mass spectral data on the corresponding tri-N-acetyl-O-trimethylsilyl derivative, which was prepared according to a standard procedure¹⁴; calcd. for a C₃₀H₆₃O₁₀N₃Si₄ fragment (deoxystreptamine, glucosamine portions), 737.3590; found 737.3586; calcd. for a C₂₇H₅₇O₈N₂Si₄ fragment (deoxystreptamine, ribose portion), 649.3192; found 649.3198.

It is noteworthy to mention that in the synthesis of ribostamycin, Ito and coworkers¹⁵ effected a similar glycosidation reaction between the halide 3 and tri-N-benzyloxycarbonyl 3',4'-di-O-benzyl neamine, and reported the isolation of a major product in which the D-ribofuranosyl moiety was attached to the 5-position of the deoxystreptamine unit. It is somewhat surprising that glycosidation under essentially similar conditions, and of the same unit in the structurally related paromamine and neamine derivatives, respectively, leads to such widely divergent substitution patterns¹⁶.

The pseudotrisaccharide 6 was much less active against bacteria, compared to its positional isomer described in the accompanying paper⁶. These results will be communicated at a later date.

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